THE MICROPLATE AGAR - REPLACEMENT, REDUCTION AND REFINEMENT OF THE MICROSPUMENSION SALMONELLA/MICROSOME ASSAY


Abstract
The Salmonella/microsome assay is the most used mutagenicity test both for evaluation of chemicals and environmental samples. The objective of this study was to miniaturize the microspumension version of this assay under the concept of the 3R.

Key words:
Mutagenicity, 3R, Ames.

Introduction
The Salmonella/microsome assay is the most used mutagenicity test both for evaluation of chemicals and environmental samples1. There are several versions of protocols available in the literature2-4. Miniaturization of toxicological tests has been a tendency in compliance with the concept of the 3Rs (Replacement, Reduction and Refinement). The microspumension version of this assay is performed with conventional petri dishes (90 x 15 mm) and uses 5X concentrated bacteria and 10 times less sample and S9 mixture. It has been extensively used for environmental samples testing, including in Effect Directed Analysis (EDA). It can be used with diagnostic strains, which have different spontaneous reversion rates. The objective of this study was to miniaturize of the microspumension Salmonella/microsome assay using agar microplates under the concept of the 3R.

Results and Discussion
Following the same principles of the microspumension protocol and aiming a reduction of the assay in 10 times. The conventional plates were replaced by microplates with 12 wells, and the method which is illustrated in the Image 1 was refined to correct the problem of using small amounts of sample (less then 2 µL). For validation of the miniaturization, three strains (TA1538, TA98 and YG1041) were selected because of their different spontaneous reversion rates (low, mean and high), and tested with mutagenic chemicals. The microspumension and miniaturization procedures were made as similar as possible conditions possible, and similar results were obtained when compared (see Chart 2). The advantages of using the miniaturization are presented in the Chart 1.

Conclusions
MPA and Microspumension provided similar sensitivity. The MPA is less laborious, uses less sample, materials and reagents. MPA procedure seems to be a promising tool specially to test environmental samples for mutagenic activity when quantity of sample is a limiting factor.

Acknowledgement
The authors thank FAPESP Project 2013/16956-6. José Ricardo R. M. Zwarg thanks FAPESP Project 2015/11399-7 for the IC scholarship. Daniel A. Morales thanks CAPES for the PhD scholarship. The SOLUTIONS project has received funding from the European Unions Seventh Framework Programme for research, technological development and demonstration under grant agreement no. 603437.

References


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**Chart 1.** Comparison of the MPA and Microspumension protocols (Kado) for TA1538, TA98 and YG1041 strains of Salmonella typhimurium in the presence (+) and absence (-) of metabolic activation (S9).

<table>
<thead>
<tr>
<th>Sample</th>
<th>TA1538</th>
<th>TA98</th>
<th>YG1041</th>
</tr>
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<tbody>
<tr>
<td>S9</td>
<td>MPA</td>
<td>Kado</td>
<td>MPA</td>
</tr>
<tr>
<td>1NTP</td>
<td>0.016</td>
<td>0.013</td>
<td>0.0015</td>
</tr>
<tr>
<td>BaP</td>
<td>2.0</td>
<td>1.6</td>
<td>3.5</td>
</tr>
<tr>
<td>4NQO</td>
<td>0.03</td>
<td>0.03</td>
<td>0.03</td>
</tr>
</tbody>
</table>

**Chart 2.** Comparison of the MPA and Microspumension protocols (Kado) for TA1538, TA98 and YG1041 strains of Salmonella typhimurium in the presence (+) and absence (-) of metabolic activation (S9).